

Efficacy of *Nigella Sativa* (Ranunculaceae) Extracts on Adult Performance and Phase Transition of the Desert Locust *Schistocerca gregaria* (Orthoptera: Acrididae)

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Abstract: The current work was carried out to investigate the effects of methanolic, petroleum ether and n-butanol extracts (30.0, 15.0, 7.5, 3.7 and 1.8%) of *Nigella sativa* seeds on several parameters of the adult performance and phase transition of *Schistocerca gregaria*. The n-butanol extract exhibited the most potent adulticidal activity followed with petroleum ether and methanolic extract, respectively, after treatment of penultimate (4th) instar nymphs. After treatment of last (5th) instar nymphs, methanolic extract exhibited the least adulticidal activity. Also, treatment of penultimate instar nymphs with *N. sativa* extracts resulted in blocked adult emergence in a dose-dependent course. Whereas no effect was exhibited by n-butanol extract on adult emergence after treatment of last instar nymphs, various degrees of restrained process was determined at some concentrations of other extracts. All *N. sativa* extracts (only at the higher two concentrations) caused adult deformities after treatment of the penultimate instar nymphs. After treatment of the last instar nymphs, n-butanol extract halted the adult morphogenesis only at the higher two concentrations but other extracts impaired it at all concentrations. In connection with the phase transition, treatment of penultimate instar nymphs with n-butanol extract (at 15.0 %) resulted in a solitarious tendency of *S. gregaria* adults as appeared with deeply green colour. The ovarian maturation in adult females was pronouncedly or slightly prohibited by *N. sativa* extracts during prolonged duration, depending on the concentration. Also, the reproductive life-time (oviposition period) was affected. Total adult longevity was shortened or prolonged, i.e. adult aging was accelerated or delayed, depending on extracts, concentration level and time of treatment.

Keywords: Emergence, Longevity, Methanol, Morphogenesis, Mortality, N-Butanol, Petroleum Ether, Solitarization.

1. INTRODUCTION

The desert locust *Schistocerca gregaria* (Forsk.) (Orthoptera: Acrididae) ranks together with other migratory locusts-amongst the most important crop pests in Africa. Damage caused by the desert locust is a consequence of its polyphagous behaviour, high density of the population, and the nature to aggregate and swarm. Each individual gregarious locust is able to consume roughly its own weight (about 2 grams) in foliage daily [1-3]. In the last century alone, there were seven periods of numerous plagues, the longest of which lasted intermittently for 13 years [2]. Current locust control operations are mainly based on organophosphorus pesticides as a result of the banning of organochlorines [4]. The widespread use of such synthetic pesticides has considerable drawbacks, such as the development of insect resistance to insecticides, increased costs, handling hazards, concerns about insecticide residues, and great threats to both human and environmental health [5]. Therefore, many institutions have intensified their efforts in the search for integrated locust control measures. Much attention has been devoted to use plant extracts or plant constituents that have insecticidal effects [6-8] because they are generally pest-specific, relatively harmless to non-target organisms and they are biodegradable and consequently harmless to the environment [9,10].

Nigella plants are widely distributed in countries which border the Mediterranean Sea, central Europe and western Asia [11]. There are many species classified in the genus *Nigella* (Ranunculaceae) [12,13]. Among the most important medicinal crops in Egypt is *Nigella sativa* which is commonly called as known as black seed or black cumin [14] and "Habbat al-barakah" (the seed of blessing) in Arabic. Seeds of *N. sativa* and their oil have a long history of folklore usage in various systems of medicines. [15] reviewed the medicinal, pharmacological, traditional value and folk remedies of this herb. In pest control, [16] reported that oleic and linoleic acid as insecticidal components from *N.*

sativa which were found to be toxic to *Callosobruchus chinensis*. Similar results were obtained [17,18]. The *N. sativa* extracts exhibited toxic effects on *Spodoptera littoralis* [19] and *S. gregaria* [20] in addition to disrupted growth, development [20] and larval haemogram [21] of the latter insect. Also, [22] studied the insecticidal activity of *N. sativa* extracts against the larvae of *Trogoderma granarium* under laboratory conditions. Recently, [23] reported disturbing effects of the acetone seed extract on biology and invasion of the stored product pest *Tribolium castaneum*. The present work was carried out to investigate the effects of different extracts of *N. sativa* on the adult performance of *S. gregaria* including emergence, survival, morphogenesis and longevity. In addition, possible effect of the present plant extracts on phase transition of *S. gregaria* was studied.

2. MATERIALS AND METHODS

2.1. Experimental Insect

The desert locust *S. gregaria* was used as an experimental insect in the present study. The insects were reared and handled under the crowded conditions of [24]. Depending on the improvements of [25] insects were reared in wooden formed cages provided with electric bulbs (150 watt) adjusted to a photoperiod of 12L:12D and to maintain an ambient temperature of $32\pm 2^{\circ}\text{C}$. Fresh clean leaves of *Trifolium alexandrinum* (Egyptian clover), in winter, and the leaves of leguminous plant *Sesbania aegyptiaca*, in summer, were used for feeding insects in the stock culture. On the other hand, *T. alexandrinum* leaves only were offered as food for insects of the experimental work.

2.2. Plant Extracts

Samples of *N. sativa* seeds were purchased from an Egyptian market. The samples were air-dried, powdered and kept in tightly closed amber coloured glass containers for protecting from light, at low temperature. Dried and pulverized powder of *N. sativa* (2 kg) was exhaustively separately extracted with methanol (1.7 Lx3). The combined alcohol extracts were concentrated to 400 ml, diluted with 400 ml of water and the next successively extracted with petroleum ether (5x400 ml) was concentrated to dryness under reduced pressure giving (11 and 90 g), and n-butanol (5x400 ml) extracts were concentrated to dryness under reduced pressure giving (75 and 55 g).

2.3. Nymphal Treatments

The newly moulted 4th (penultimate), or 5th (last) instar nymphs of *S. gregaria* were fed on fresh leaves of *Trifolium alexandrinum* after dipping in the different concentration levels of each *N. sativa* seed extract. After dipping for three minutes, the treated leaves were allowed to dry before offering to nymphs. A day after treatment, all nymphs (treated and control) were provided with untreated fresh food plant. Ten replicates (one nymph/replicate) were used for each concentration. Each individual nymph was isolated in a glass vial provided with a thin layer of sterilized sand as a floor. All vials were located in a large cage having a suitable electric bulb. The nymphs were carefully handled until the adult emergence just after which all parameters of adult performance and solitarization tendency were recorded.

2.4. Adult Performance Parameters

Adult emergence was recorded in percentage. For investigation the adulticidal activity of *N. sativa* extracts on *S. gregaria*, the adult mortality was observed throughout the adult longevity and calculated in percentage. For investigating the morphogenic efficiency, the adult deformities were observed and calculated in percentage according to [26] as follows:

$$[\text{No. of deformed adults} / \text{No. of larvae}] \times 100$$

The ovarian maturation period, reproductive life-time, post-oviposition period and total adult longevity was measured in days \pm SD [27].

The solitarization tendency of the adults appeared with green colour and other solitary features. The phase transition was estimated in percentage.

2.5. Statistical Analysis of Data

Data obtained were analyzed by the Student's *t*-distribution, and refined by Bessel correction [28] for the test significance of difference between means.

3. RESULTS

3.1. Effect of *N. Sativa* Extracts on Adult Survival

Depending on data assorted in Table (1), the survival potential of adult *S. gregaria* was affected by a latent adulticidal activity of *N. sativa* seed extracts. Treatment of penultimate (4th) instar nymphs with the highest concentration of methanolic extract resulted in 20% mortality. The same extract, at other concentrations, failed to cause adult mortality. Both petroleum ether and n-butanol extracts exhibited various adulticidal activities since different mortality percentages were recorded, regardless the concentration. Moreover, n-butanol extract was the most effective on the adult survival followed with petroleum ether and methanolic extracts, respectively. As clearly seen in the same table, a similar adulticidal activity of *N. sativa* extracts could be exhibited after treatment of last instar nymphs. Furthermore, mortality was dose-dependent by both petroleum ether and n-butanol extracts. Methanolic extract was the least toxic one (10.0% mortality at the highest concentration vs. 0.0% mortality of control adults).

3.2. Effect of *N. Sativa* Extracts on Adult Emergence

Data of Table (2) clearly reveal some effects of *N. sativa* extracts on the nymphal metamorphosis into adults after treatment of penultimate instar nymphs because the adult emergence decreased as the concentration was increased. As for example, the adult emergence was determined as 62.5 and 20.0 (compared to 88.9% of control congeners) at the highest concentration of methanolic and petroleum ether extracts, respectively. No adults emerged after treatment with the highest concentration of n-butanol extract but the sublethal concentration led to only 50.0% of adult emergence (compared to 90.0% of control congeners). Whereas no effect was displayed by n-butanol extract on the adult emergence after treatment of last instar nymphs, restrained emergence was observed after treatment with the higher two concentrations of petroleum ether extract (60 and 40%, respectively, vs. 90% emergence of control adults). Also, treatment with methanolic extract, at 30.0 and 3.7%, resulted in 90.0% adult emergence (compared to 100% emergence of adult controls, Table 2).

Table1. Adulticidal activity (%) of *N. sativa* extracts on *S. gregaria*

Solvent	Conc. (%)	After treatment of 4th instar nymphs	After treatment of 5th instar nymph
Methano	30.0	20.0	10.0
	15.0	00.0	00.0
	07.5	00.0	00.0
	03.7	00.0	10.0
	01.8	00.0	00.0
	Controls	00.0	00.0
Petroleum ether	30.0	50.0	50.0
	15.0	12.5	50.0
	07.5	14.2	22.2
	03.7	14.2	22.2
	01.8	00.0	11.1
	Controls	00.0	00.0
n-butanol	30.0	---	22.2
	15.0	50.0	22.2
	07.5	50.0	42.9
	03.7	40.0	22.2
	01.8	33.3	11.1
	Controls	00.0	00.0

Conc.: Concentration level. ---: No adult could metamorphose from the treated nymphs

3.3. Effect of *N. Sativa* Extracts on Adult Morphogenesis

In connection with the impaired adult morphogenesis program of *S. gregaria* by *N. sativa* seed extracts, data distributed in the previously cited table obviously show various percentages of deformed adults. After treatment of the penultimate instar nymphs, only 20% adult deformities were recorded at the highest concentration of methanolic extract but it failed to affect the morphogenesis at other lower concentrations.

Table 2. Affected adult emergence and morphogenesis of *S. gregaria* by nymphal treatments with *N. sativa* extracts

Solvent	Conc. (%)	After treatment of 4th instar nymphs			After treatment of 5th instar nymphs		
		Emergence (%)	Deformed %	Solitarian %	Emergence (%)	Deformed %	Solitarian %
Methanol	30.0	62.5	20.0	0.0	090.0	0.0	0.0
	15.0	85.7	0.0	0.0	100.0	0.0	0.0
	07.5	88.9	0.0	0.0	100.0	0.0	0.0
	03.7	87.5	0.0	0.0	090.0	0.0	0.0
	01.8	88.9	0.0	0.0	100.0	0.0	0.0
	Controls	88.9	0.0	0.0	100.0	0.0	0.0
Petroleum ether	30.0	20.0	50.0	0.0	60.0	33.3	0.0
	15.0	85.7	12.5	0.0	40.0	25.0	0.0
	07.5	85.7	14.2	0.0	90.0	11.1	0.0
	03.7	85.7	14.2	0.0	90.0	22.2	0.0
	01.8	88.9	0.0	0.0	90.0	11.1	0.0
	Controls	88.9	0.0	0.0	90.0	0.0	0.0
n-butanol	30.0	---	---	0.0	90.0	22.2	0.0
	15.0	50.0	40.0	50.0	90.0	11.1	0.0
	07.5	40.0	33.3	0.0	90.0	0.0	0.0
	03.7	60.0	33.3	0.0	90.0	11.1	0.0
	01.8	60.0	25.0	0.0	90.0	11.1	0.0
	Controls	90.0	0.0	0.0	90.0	0.0	0.0

Conc., ---: see footnote of Table (1). Mean \pm SD followed by letter (a): not significantly different ($P>0.05$), (b): Significantly different ($P<0.05$), (c): Highly significantly different ($P<0.01$), (d): Very highly significantly different ($P<0.001$).

Petroleum ether and n-butanol extracts were more potent because different percentages of adult malformations were observed almost proportionally to the concentration (50.0, 14.2, 14.2 and 12.5 at 30.0, 15.0, 7.5 and 3.7 % of petroleum ether extract as well as 40.0, 33.3, 33.3 and 25.0 at 15.0, 7.5, 3.7 and 1.8 of n-butanol extract). The adult deformities could be, generally, assorted in the following features: Adults with curled legs and coiled incompletely developed short antennae. Adults with crumpled wings and transparent posterior area and coiled antennae (Fig.1).



Fig1. Different adult malformations of *S. gregaria* were produced as a result of the nymphal treatments with *N. sativa* extracts. A) Normal adult. B) Treated adult with curled legs, incompletely developed short antennae and crumpled wings with transparent posterior area. C) Treated adult with crumpled wings of transparent posterior area and coiled antenna.

Adult failure to completely get rid the last nymphal exuvia, where the nymphal exuvia remained as attached parts to the adult body (Fig. 2). After treatment of the last instar nymphs with methanolic extract, no deranging action could be exerted on the adult morphogenesis (Table 2). In contrast, treatment with petroleum ether or n-butanol extracts resulted in serious adult deformities. At the highest concentration of each, the strongest action was exerted on morphogenesis (33.3 % adult

deformities at 30 % of petroleum ether extract and 22.2 % adult deformities at 30 % of n-butanol extract (compared to no adult deformities of control adults). Referring to Figs 1 and 2, features of adult impaired morphogenesis program can be observed and described as previously mentioned.



Fig2. Different degrees of adult failure to completely get rid the last nymphal exuvia as a result of the nymphal treatments with *N. sativa* extracts. A) Nymphal exuvium attached to abdomen, wings and legs. B) Nymphal exuvium attached to wings, legs and mouth parts. C) Nymphal exuvium attached to wings.

3.4. Effects of *N. Sativa* Extracts on Phase Transition

After treatment of penultimate instar nymphs with only n-butanol extract of *N. sativa*, an important solitarization affect was exhibited because 50 % of the deformed adults appeared with some characteristics of the solitary phase (such as deeply green colour of the body) at 15 % of n-butanol extract (Table 2 and Fig. 3). No solitarization effect was recorded after treatment of last instar nymphs, regardless the extract or concentration.

3.5. Effects of *N. Sativa* Extracts on Adult Longevity

It may be conceivable to mention that the maturation period (preoviposition period) is an important indicator for the ovarian maturation rate, i.e, longer period usually indicate a slower rate and *vice versa*. After treatment of penultimate instar nymphs with *N. sativa* seed extracts, data arranged in Table (3) exiguously show that methanolic extract pronouncedly prohibited the ovarian maturation of *S. gregaria* during remarkably lengthened duration, especially at the higher three concentrations (31.3 ± 1.5 , 28.7 ± 1.5 and 28.7 ± 0.6 days at 30.0, 15.0 and 7.5 % vs. 22.0 ± 1.7 days of controls). On the other hand, both petroleum ether and n-butanol extracts slightly prohibited it, during insignificantly prolonged duration, regardless the concentration.

After treatment of last instar nymphs, data of the same table clearly indicate a major prolonging effect of *N. sativa* seed extracts on the ovarian maturation period which may be informative to delayed sexual maturity owing to regressed ovarian maturation rate, especially at the higher concentrations. However, methanolic extract pronouncedly prohibited such vital process at the higher three concentrations (25.7 ± 1.1 , 26.0 ± 1.3 and 26.0 ± 1.0 days, at 30.0, 15.0 and 7.5%, vs. 22.0 ± 1.7 days of control congeners). Only at the higher two concentrations of petroleum ether extract and the highest concentration of n-butanol extract, the ovarian maturation period was significantly prolonged indicating remarkably delayed sexual maturity (Table 3).

Considering the reproductive life-time (oviposition period), data assorted in Table 4 show general enforcing action of *N. sativa* extracts on the adult females to quickly lay eggs during shortened period, after treatment of penultimate instar nymphs. Such action was exerted during significantly or insignificantly shortened period, depending on the concentration of methanolic extract and petroleum ether extract. Moreover, n-butanol extract exerted stronger enforcing action on this process at the majority of concentrations (at least $P < 0.05$: 7.7 ± 1.5 , 8.7 ± 0.6 , 9.7 ± 1.5 and 11.7 ± 1.2 days at 15.0, 7.5, 3.7 and 1.8 %, compared to 13.7 ± 2.1 days of controls). After treatment of last instar nymphs, a prohibiting effect was appreciated for adult females by methanolic extract because they lasted

insignificantly prolonged reproductive life time. A reverse result was recorded for both petroleum ether and n-butanol extracts because adult females had been enhanced to lay eggs during shortened time intervals (11.7 ± 1.6 and 11.3 ± 1.5 , $p < 0.01$, at 30.0 and 15.0 % of petroleum ether extract, vs. 23.7 ± 1.2 days of controls, as well as 9.3 ± 1.0 and 11.3 ± 1.5 , $p < 0.05$ at least, at 30.0 and 15.0 % of n-butanol extract, vs. 15.7 ± 1.5 days of controls, Table 4).

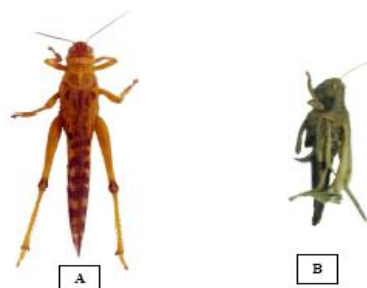


Fig3. Phase shift of *S. gregaria* from gregarious to solitaria as a result of the nymphal treatments with some concentrations of *N. sativa* extracts. A) Normal gregarious adult. B) Solitarized adult.

Table3. Influenced ovarian maturation period (Mean days \pm SD) of *S. gregaria* by nymphal treatments with *N. sativa* extracts.

Solvent	Conc. (%)	After treatment of 4th instar nymph	After treatment of 5th instar nymphs
Methanol	30.0	31.3 ± 1.5 c	25.7 ± 1.1 b
	15.0	28.7 ± 1.5 c	26.0 ± 1.3 b
	07.5	28.7 ± 0.6 c	26.0 ± 1.0 b
	03.7	24.0 ± 1.7 a	24.3 ± 1.2 a
	01.8	23.7 ± 1.5 a	24.0 ± 1.0 a
	Controls	22.0 ± 1.7	22.0 ± 1.7
Petroleum ether	30.0	---	28.3 ± 1.2 c
	15.0	25.0 ± 1.0 a	26.7 ± 1.3 b
	07.5	23.0 ± 1.7 a	24.7 ± 1.5 a
	03.7	23.0 ± 1.0 a	24.3 ± 0.6 a
	01.8	22.7 ± 1.2 a	24.7 ± 1.2 a
	Controls	22.0 ± 1.7	23.7 ± 1.2
n-butanol	30.0	---	24.0 ± 1.0 c
	15.0	27.7 ± 3.1 a	28.3 ± 1.2 a
	07.5	28.7 ± 4.2 a	27.7 ± 1.2 a
	03.7	28.0 ± 3.6 a	27.7 ± 0.6 a
	01.8	30.7 ± 1.5 a	28.3 ± 0.6 a
	Controls	32.0 ± 3.7	28.0 ± 1.0

Conc., ---: see footnote of Table (1). a, b, c, d: see footnote of Table (2).

Table4. Disturbed reproductive life-time (Mean days \pm SD) of *S. gregaria* by nymphal treatments with *N. sativa* extracts

Solvent	Conc. (%)	After treatment of 4th instar nymphs	After treatment of 5th instar nymphs
Methanol	30.0	10.7 ± 1.2 c	17.3 ± 1.2 a
	15.0	14.0 ± 1.7 a	17.0 ± 1.0 a
	07.5	14.0 ± 1.0 a	16.7 ± 1.2 a
	03.7	16.7 ± 1.2 a	17.3 ± 2.1 a
	01.8	16.3 ± 1.5 a	17.3 ± 1.5 a
	Controls	16.7 ± 1.5	16.7 ± 1.5
Petroleum ether	30.0	---	11.7 ± 1.6 c
	15.0	08.7 ± 1.2 c	11.3 ± 1.5 c
	07.5	09.7 ± 1.5 c	17.0 ± 1.0 a
	03.7	15.3 ± 1.5 a	17.7 ± 1.2 a

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n-butanol	01.8	16.3 ± 1.5 a	17.0 ± 2.0 a
	Controls	16.7 ± 1.5	19.0 ± 1.0
	30.0	---	09.3 ± 1.2 c
	15.0	07.7 ± 1.5 c	11.3 ± 1.5 b
	07.5	08.7 ± 0.6 c	13.3 ± 1.5 a
	03.7	09.7 ± 1.5 b	14.7 ± 1.2 a
	01.8	11.7 ± 1.2 a	15.3 ± 0.6 a
	Controls	13.7 ± 2.1	15.7 ± 1.5

Conc., ---: see footnote of Table (1). a, b, c, d: see footnote of Table (2).

The total adult longevity can be used as an informative indicator of the adult aging, i.e. the prolonged longevity denotes the delaying of adult aging and vice versa. Data of total adult longevity, as affected by the *N. sativa* extracts, were listed in Table (5). After treatment of penultimate instar nymphs, both methanolic and petroleum ether extracts caused a slight prolongation in the total longevity, irrespective of concentration. In contrast, n-butanol extract exhibited a pronounced shortening effect on longevity because all treated adult females reached the death point after remarkably shorter duration than that of control adult females, at all concentrations (39.7±3.5, 43.0±3.6, 43.0±3.5 and 48.7±2.5 days at concentrations 15.0, 7.5, 3.7 and 1.8 %, vs. 58.8±4.6 days of controls).

After treatment of the last instar nymphs, data of aforementioned table obviously revealed a shortening effect of both petroleum ether and n-butanol extracts on the total longevity which was obviously observed at the higher two concentrations (30.0 and 15.0 %, respectively). In other words, petroleum ether and n-butanol extracts led to an accelerated aging of the adults ending in death (45.0±1.0 and 45.7±3.1 days, compared to 53.0±2.6 days of controls, for petroleum ether extract and 38.7±2.3 and 48.0±2.6 days, compared to 53.0±1.0 days of controls, for n-butanol extract). On the contrary, methanolic extract did not exert a similar action but reversely delayed the adult aging during slightly prolonged longevity.

Table5. Disturbed total adult longevity (Mean days±SD) of *S. gregaria* by nymphal treatments with *N. sativa* extracts

Solvent	Conc. (%)	After treatment of 4th instar nymphs	After treatment of 5th instar nymphs
Methanol	30.0	46.3 ± 2.5 a	45.7 ± 2.1 a
	15.0	46.3 ± 1.5 a	45.7 ± 2.9 a
	07.5	46.7 ± 1.2 a	45.3 ± 1.5 a
	03.7	44.7 ± 3.5 a	45.0 ± 2.0 a
	01.8	44.3 ± 1.5 a	45.0 ± 1.0 a
	Controls	43.3 ± 2.1	43.3 ± 2.1
Petroleum ether	30.0	---	45.0 ± 1.0 c
	15.0	42.3 ± 2.1 a	45.7 ± 3.1 b
	07.5	41.3 ± 3.5 a	48.3 ± 1.5 a
	03.7	43.3 ± 3.2 a	49.3 ± 1.5 a
	01.8	44.0 ± 1.0 a	49.0 ± 3.5 a
	Controls	43.3 ± 2.1	53.0 ± 2.6
n-butanol	30.0	---	38.7 ± 2.3 d
	15.0	39.7 ± 3.5 c	48.0 ± 2.6 b
	07.5	43.0 ± 3.6 c	49.0 ± 2.6 a
	03.7	43.0 ± 3.5 c	50.7 ± 1.5 a
	01.8	48.7 ± 2.5 b	52.3 ± 1.2 a
	Controls	58.8 ± 4.6	53.0 ± 1.0

Conc., ---: see footnote of Table (1). a, b, c, d: see footnote of Table (2).

4. DISCUSSION

4.1. Blocked Adult Emergence of *S. gregaria*

Complete or partial blockage of adult emergence was reported for different insects by various botanicals such as the blocked emergence of *Musca domestica* [29] and *Rhynchophorus ferrugineus* by azadirachtin [30], *Tribolium castaneum* by the methanolic extracts of *Centaurium erythrae* and

Pteridium aquilinum [31], *S. gregaria* by extracts of *Fagonia bruguieri* [32] and *Ammi visnaga* [33] as well as *Earias vittella* by Neemazal T/S and Nimbecidine [34].

In the present study, treatment of penultimate (4th) instar nymphs of *S. gregaria* with *N. sativa* seed extracts resulted in blocked adult emergence in a dose-dependent course. Whereas no effect was exhibited by n-butanol extract on adult emergence after treatment of last (5th) instar nymphs, various degrees of restrained emergence was determined at the higher two concentrations of petroleum ether extract (30.0, 15.0%) and at 30.0 and 3.7% of methanolic extract. Since the eclosion hormone, a blood-born factor arising from the central nervous system [35] triggers eclosion in a wide range of insect orders including Orthoptera [36], the *N. sativa* extracts probably prevented this hormone from being released at the appropriate time. Hence, the eclosion hormone appears to be affected by a certain active ingredient(s) contained in the *N. sativa* extracts. However, the exact mode of action needs further investigation.

4.2. Affected Adult Survival of *S. gregaria*

The available literature contains many reported toxicities of extracts from various plant species on the immature stages of several insect pests [37, 38, 39, 25, 33; 40, 41, 42, 43, 44, 45, 46, 47] while the lethal effects of botanicals on adults are relatively scarce. In the present study, n-butanol extract of *N. sativa* seeds exhibited the most potent adulticidal activity followed with petroleum ether and methanolic extract, respectively, after treatment of penultimate instar nymphs of *S. gregaria*. The methanolic extract exhibited the least mortal effect after treatment of last instar nymphs. These results agree, to some extent, with those reported adulticidal activities of different plant species on some pests, such as *T. castaneum* [48], *Muscina stabulans* [49] and *M. domestica* [50]. Also, the current results are in consistent with the adulticidal activities of extracts derived from *Rhizophora mucronata* [51], *Fagonia bruguieri* [32] and *Punica granatum* [52] on the same locust.

The adult mortality, i.e., reduced survival potential, of *S. gregaria* by *N. sativa* extracts, in the present study, may be explicated by a latent prohibitory effect on feeding leading to continuous starvation and subsequently death [39]. It may be, also, attributed to the action of certain active ingredients in the *N. sativa* seed extracts on the homeostasis leading to increasing loss of body water and subsequently death [50], since *N. sativa* contains conjugated linoleic acid, thymoquinone, nigellone (dithymoquinone), melanthin, nigilline, damascenine, tannins, flavonoids, saponins, alkaloids, proteins, lipids, dithymoquinone carvacol and anethole 4-terpinole [53, 54, 55, 15, 56].

4.3. Deranged Adult Morphogenesis of *S. gregaria*

In the present work, all *N. sativa* extracts (only at the higher two concentrations) caused adult deformities after treatment of the penultimate instar nymphs. After treatment of the last instar nymphs, n-butanol extract halted the adult morphogenesis only at the higher two concentrations but other extracts impaired it at all concentrations. These results are in agreement with those reported results for extracts from various plants against the same locust. As for examples, adult morphogenic defects were observed after treatment of last instar nymphs with a neem oil [57], after treatment of penultimate instar nymphs with ethanol extract of *Cyprus rotendus* [58], Neemazal (a neem preparation) [20], some extracts of *F. bruguieri* [32] as well as some extracts of *P. granatum* peel [52]. Moreover, various malformed moths of *Spodoptera littoralis* were caused by Neemazal [39], acetone and ethanol extracts of *Aristolochia pubescens* impaired the adult morphogenesis of *Aticarsia gemmatalis* [59], as well as many adult deformities in both *Spodoptera frugiperda* and *Tenebrio molitor* were observed after treatment with methanol extract of *Myrtillocactus geometrizans* [60].

Imperfectly emerged adults, in the present study may be due to the disturbance of normal ecdysteroid titer which is usually needed for the achievement of perfect metamorphosis program or even the inhibition of neurosecretion (prothoracicotropic hormone) causing inhibition of a number of physiological processes, such as metamorphosis and morphogenesis [61].

4.4. Induced Solitarization Tendency of *S. gregaria*

The desert locust, *S. gregaria*, usually display a dramatic polyphenism, being able to transform reversibly between two forms or phases that differ considerably in many aspects including behaviour, physiology and morphology [62-68]. Many studies have been performed searching for the exogenous and endogenous causes of phase changes in *S. gregaria*. They has focused on the changes from gregarious to solitary, since only gregarious locusts form large migratory swarms capable of invading

and inflicting serious damage to crops. No striking interpretation was introduced more than the suggestion about the role of ecdysteroids, juvenoids, and possibly also pheromones in initiating and regulating this process [69].

As reported in the available literature, phase shift from gregaria to solitaria in *S. gregaria* was caused by some extracts of *Melia volkensii* [70,71]. A clear tendency to solitarization was elicited after treatment of *S. gregaria* gregarious phase with neem oil (37, 57, 72). Also, treatment of earlier instar nymphs of *Locusta migratoria migratorioides* resulted in behavior toward the solitary phase [73]. Treatment of gregarious penultimate or last instar nymphs of *S. gregaria* with the ethanol extract of *C. rotendus* resulted in a solitary tendency in adult females [74]. The n-butanol extract of *F. bruguieri* enhanced the solitarious tendency in adult females of gregarious *S. gregaria*, regardless the time of nymphal treatment [32]. In connection with the phase transition, in the present study, treatment of penultimate instar nymphs of *S. gregaria* with n-butanol extract of *N. sativa* seeds (at 15.0 %) induced the solitarious tendency of *S. gregaria* because 50 % of the deformed adults appeared with deeply green colour (characteristic of solitary phase).

The phase transition can be explained on the hormone basis. Allatectomy (surgical removal of corpora allata, CA, responsible for the production of juvenile hormone, JH) resulted in no gregarious behavior in locusts [75]. Such observation rationally explains the higher activity of CA in solitary *S. gregaria* causing higher titers of JH in haemolymph and a green colouration of the cuticle [76]. On the pheromone basis, the existence of 'gregarization pheromone' was postulated [77,78]. The solitarization effect of *N. sativa* n-butanol extract, in the present study, may be due to their influence on this pheromone or to its influence on the hormonal system of the insect [72]. For some detail, JH influences the response of olfactory interneurons in the antennal lobe to aggregation pheromone, whereas the responsiveness of antennal receptors neurons is not changed [75]. In conclusion, it is reasonable to suggest the existence of a juvenilizing, and subsequently antigregarizing, substance in *N. sativa* extracts but more deep investigation is needed to disclose some aspects of our suggestion since juvenilizing effects of some other plant species, such as *Ajuga chamaepitys*, were determined [79].

4.5. Disturbed Adult Longevity of *S. gregaria*

In Orthoptera, the sexual maturity usually needs a time interval elapsed between adult emergence until the day of laying the first egg. During such period, the ovaries (or testes) developed and the adult will be sexually mature. Generally, the pre-oviposition period may be informative for the sexual maturity rate, i.e. the shorter period indicates the faster rate and *vice versa*. Thus, it may acceptable to use the pre-oviposition period in adult females of *S. gregaria* as a good indicator to the ovarian maturation rate. In this regard, several contradictory results had been reported in the literature, since some plant extracts promoted the ovarian maturation, and hastened the sexual maturity, while others prohibited the ovarian maturation, and delayed the sexual maturity. An enhancing effect on the ovarian maturation of *S. gregaria* was exhibited by certain concentrations of Neemazal (a neem preparation) [20] as well as by methanolic and petroleum ether extracts of *F. bruguieri* [32]. In contrast, some extracts of *C. rotendus* completely retarded the ovarian maturation of the same locust [58], n-butanol extract of *F. bruguieri* exhibited a delaying effect on the same process [32] and some extracts of *P. granatum* peel slightly or remarkably retarded this vital process [52]. On the other hand, no effect was exhibited on it in *M. domestica* by Margosan-O (a neem preparation) or Jojoba oil [80].

In the present investigation, treatment of penultimate instar nymphs of *S. gregaria* with methanolic extract of *N. sativa* seeds resulted in pronouncedly prohibited ovarian maturation but petroleum ether extract or n-butanol extract exhibited a slight inhibitory effect. Moreover, predominantly retarding effect on this vital process during prolonged duration was recorded after treatment of last instar nymphs, especially at the higher concentrations. An appreciable interpretation of the prolonged pre-oviposition period, indicating delayed sexual maturity and regressed ovarian maturation rate, in *S. gregaria* after treatment with *N. sativa* extracts, in the present study, is still obscure but some active compounds in these extracts may interfere with the hormonal regulation of this physiological event.

As reported in the literature, treatments of some insects with extracts of various plants resulted in shortened reproductive life-time (oviposition period) of the adult females. With regard to *S. gregaria*, treatment of 2nd-4th instar nymphs with ethanol extract of *M. volkensii* shortened the reproductive life-time [71]. A similar result was reported after nymphal treatments with *F. bruguieri* [81] or *P.*

granatum peel extracts [52]. In addition, shortened reproductive life-time of some other insects was caused by several botanicals, such as *M. domestica* by an aqueous extract of *Hyoscyamus muticus* [82] and Margosan-O or Jojoba oil [81] and *Chrysomya chloropyga* by some extracts of certain Nigerian plants [83]. In agreement with these reported results, the current study revealed an enforcing action of all *N. sativa* seed extracts on the reproducing adult females of *S. gregaria* to quickly lay eggs during significantly or insignificantly shortened period. An exceptional case of prolonged time was recorded after treatment of last instar nymphs only with methanolic extract. Unfortunately, no acceptable interpretation of the general shortening effect of *N. sativa* extracts on the reproductive life-time, or enforcing the adult females of *S. gregaria* to quickly lay eggs, is available right now!! Therefore, further investigation should be carried out to explore the mode of action of certain chemical constituents of these extracts on this crucial physiological criterion.

After the attainment of sexual maturity, insects often show degenerative changes in some tissues and organs which can be called 'senility' or 'aging'. In insects, the affected adult longevity can be considered an informative indicator of the adult aging, i.e. prolongation of longevity may denote a delay of aging and *vice versa*. As reported in the available literature, several neem products pronouncedly affected the total adult longevity of some insect pests, such as *Spodoptera litura* [84-88], *M. stabulans* [49], *M. domestica* [50], *Chrysomya megacephala* [44]. Also, the adult longevity of *S. littoralis* was shortened by larval treatments with extracts from *Melia azedarach* [89, 90]. Considering the present experimental locust, *S. gregaria*, Neemazal treatments of penultimate instar nymphs resulted in remarkably shortened adult longevity but a reversal effect was recorded after treatment of last instar nymphs [91]. Similar results had been reported for the same locust by extracts of *F. bruguieri* [32]. Also, accelerating or delaying action of *P. granatum* peel extracts was exhibited on the adult females of the same locust, depending on the extract and time of nymphal treatment [52].

In the present study, treatment of penultimate instar nymphs of *S. gregaria* with methanolic extract or petroleum ether extract of *N. sativa* seeds resulted in a slight prolongation of total adult longevity (delayed aging), irrespective of the concentration. On the contrary, n-butanol extract exhibited a significant shortening effect on the longevity (accelerated aging). After treatment of the last instar nymphs, a shortening effect of both petroleum ether and n-butanol extracts was remarkably exhibited on the longevity, at the higher two concentrations. In contrast, methanolic extract affected the adult life in an insignificantly prolonged longevity or delayed aging. The probable cause of shortened and prolonged adult longevity, as described by [85-87], is due to azadirachtin's interference with the neuro-endocrine system of the insects. Delaying of adult aging (or prolonged longevity) in *S. gregaria*, in the current study, may be attributed to the antioxidant properties of some constituents of *N. sativa* seeds as extracted by certain solvents. On the other hand, accelerating of adult aging (or shortened longevity) may be explicated by the action of some chemicals extracted from the tested plant by certain solvents on a hormonal activity because there is a close relation between certain hormones and adult longevity [92-95].

5. CONCLUSION

As clearly shown in the present study, *N. sativa* seed extracts exhibited slight or remarkable effects on various parameters of adult performance of *S. gregaria*. In addition, n-butanol extract induced the phase transition from gregaria to solitaria. It prohibited the gregarization tendency of *S. gregaria* and hence the swarm formation necessary for invasion can be avoided. Therefore, *N. sativa* seed extracts can be used as a complementary agent in the integrated control of this destructive locust. However, further investigation should be carried out to ascertain the active ingredient (s) contained in these extracts responsible for these effects.

REFERENCES

- [1] Youdeowei A., Major arthropod pests of food and industrial crops of Africa and their economic importance. In: "Biological control: A sustainable solution to crop pest problems in Africa". (Yaninek J.S. and Herren H.R., eds.). Proc. of the Int. Conf. and Workshop of the IITA, Cotonou, Benin, 31-50(1988).
- [2] Lindsey R., Locusts. <http://earth.Observatory.NASA.Gov/Observatory/>(2002).
- [3] Lecoq M., Desert locust management: from ecology to anthropology. *J. Orthoptera Res.* 14(2), 179-186(2005).
- [4] Lecoq M., Recent progress in desert and migratory locust management in Africa. Are preventive actions possible? *J.Orthoptera Res.* 10, 277-291(2001).

- [5] Garriga M. and Caballero J., Insights into the structure of urea-like compounds as inhibitors of the juvenile hormone epoxide hydrolase (JHEH) of the tobacco hornworm *Manduca sexta*: analysis of the binding modes and structure-activity relationships of the inhibitors by docking and CoMFA calculations. *Chemosphere* 82, 1604-1613(2011).
- [6] Schmutterer H., Properties and potential of natural pesticides from the neem tree, *Azadirachta indica*. *Ann. Rev. Entomol.* 35, 271- 297(1990).
- [7] Schmutterer H., Insektizide aus dem Niembaum *Azadirachta indica*. *Sanfte Chemie Fur den integrierten Pflanzenschutz in Entwicklungs- und industrielandern.* *Plits* 8 (2), 57-71(1990).
- [8] Krall S. and Wilps H., New trends in locust control. *Deutsche Gesellschaft für Technische Zusammenarbeit (GTZ) GmnH.* Eschborn, Germany. 182pp. (1994).
- [9] Rembold H., Secondary plant products in insect control with special reference to the azadirachtins. In "Advances in invertebrate reproduction"(Engels W.E., ed.), Vol. 3. Amsterdam: Elsevier Science Publishing Company, pp. 481-491(1984).
- [10] Isman M.B., Perspective botanical insecticides: for richer, for poorer. *Pest Manage. Sci.* 64, 8-11(2008).
- [11] Hedrick U., *Sturtevant's Edible Plants of the World.* Dover, New York, pp. 388-389(1972).
- [12] Bailey H., *A concise dictionary of plants cultivated in United States and Canada.* Macmillan Publishing Co., Inc. New York (1978).
- [13] Atta M.B., Some characteristics of *Nigella* (*Nigella sativa* L.) seed cultivated in Egypt and its lipid profile. *Food Chemistry* 83, 63-68(2003).
- [14] Rayan H.Z., Wagih H.M. and Atwa M.M., Efficacy of Black Seed oil from *Nigella sativa* against murine infection with cysts of Me49 strain of *Toxoplasma gondii*. *Parasitologists United J.* 4(2), 165-176(2011).
- [15] Sharma N.K., Ahirwar D., Jhade D. and Gupta S., Medicinal and phamacological potential of *Nigella sativa*: a review. *Ethnobotanical Rev.* 13, 946-955(2009).
- [16] Deshpande R.S., Adhikary P.R. and Tipnis H.P., Stored grain pest control agents from *Nigella sativa* and *Pogostemon heyneanus*. *Bull. Grain Technol.* 12(3), 232-234(1974).
- [17] Adebowale K.O. and Adedire C.O., Chemical composition and insecticidal properties of the underutilized *Jatropha curcas* seed oil. *Afr. J. Biotechnol.* 5(10), 901-906(2006).
- [18] Adabie-Gomez D.A., Monford K.G., Agyir-Yawson A., Owusu-Biney A. and Osaie M., Evaluation of four local plant species for insecticidal activity against *Sitophilus zeamais* Motsch. (Coleoptera: Curculionidae) and *Callosobruchus maculatus* (F) (Coleoptera: Bruchidae). *Ghana J. Agric. Sci.* 39, 147-154(2006).
- [19] Abd ELatif M.E., Abd El-Nabi L.M.A., Hussein E.H. and Abd El-Hafez Z.A., Effect of two methods of *Nigella* and *Arugula* oils extraction and its efficacy on *Spodoptera littoralis* (Boisd.). *J. Agric. Res. Kafrelsheikh Univ.* 35(4), 1069-1081(2009).
- [20] Hamadah Kh.Sh., Ghoneim K.S., El-Hela A.A., Amer S.M. and Mohammad A.A.1., Disturbed survival, growth and development of the desert locust *Schistocerca gregaria* by different extracts of *Azadirachta indica* (Meliaceae) and *Nigella sativa* (Ranunculaceae). *Egypt. Acad. J. Biolog. Sci.* 6(2), 01 -21(2013).
- [21] Ghoneim K., Hamadah Kh., Amer M., El-Hela A. and Mohammad A., Qualitative and quantitative changes in the haemogram of desert locust *Schistocerca gregaria* (Orthoptera: Acrididae) by extracts of *Nigella sativa* (Ranunculaceae). *J. Adv. Biol.* 7(2), 1275-1292(2015).
- [22] Ahmad F., Sagheer M., Hammad A., Rahman S.M.M. and Ul-Hasan M., Insecticidal activity of some plant extracts against *Trogoderma granarium* (E.). *The Agriculturists* 11(1), 103-111(2013).
- [23] Khan F.Z.A., Sagheer M., ul-Hasan M., ul-Hassan M.N., Farhan M. and Abdul Rahman, Bioactivity of *Nigella sativa*, *Syzygium aromaticum* and *Trachyspermum ammi* extracts against *Tribolium castaneum* (Herbst.) (Coleoptera: Tenebrionidae). *J. Entomol. Zool. Studies* 2(3), 103-105(2014).
- [24] Hunter-Jones P., Rearing and breeding locusts in the laboratory. *Bull. Anti-locust Res. Center London*, 12 pp. (1961).

- [25] Ghoneim K.S., Tanani M.A. and Basiouny A.L., Influenced survival and development of the desert locust *Schistocerca gregaria* (Acrididae) by the wild plant *Fagonia bruguieri* (Zygophyllaceae). Egypt. Acad. J. Biol.Sci. 2(2), 147-164(2009).
- [26] Vargas L. and Sehnal F., Use of juvenile hormone analogue against the fall webworm, *Hyphanthia cunea*. Entomol. Exp. App. 16, 115-122(1973).
- [27] Norris M.J., Sexual maturation in the desert locust, *Schistocerca gregaria* with special reference to the effect of grouping. Anti-Locust Bull. 18, 44(1954).
- [28] Moroney M.J., Facts from Figures. 3rd ed., Pinguin Book Ltd. Harmondsworth, Middlesex, 228 pp. (1956).
- [29] Naqvi S.N.H., Tabassum R., Khan M.F., Yasmin N., Nurulain S.M. and Burney A.A., Toxic, residual, and teratomorphic effect of a neem extract (N-9) in comparison to Coopex 25 WP (Permethrin + Bioallethrin) against *Musca domestica* L. (Holland strain). Turk. J. Zool. 31, 127-130(2007).
- [30] Abdel-Ghaffar A.A., Ghoneim, K.S. Tanani M.A., Bream A.S. and Nassar M.I., Developmental responses of the red palm weevil *Rhynchophorus ferrugineus* to some plant extracts. J. Egypt. Acad. Soc. Environ. Develop. 9(1), 11- 25(2008).
- [31] Jbilou R., Amri H., Bouayad N., Ghailani N., Ennabili A. and Sayah F., Insecticidal effects of extracts of seven plant species on larval development, a-amylase activity and offspring production of *Tribolium castaneum* (Herbst) (Insecta: Coleoptera: Tenebrionidae). Bioresource Technol. 99, 959-964(2008).
- [32] Aly S.A., El-Ebiarie A.S. and Hamadah Kh.Sh., Effects of the wild plant, *Fagonia bruguieri* on the adult performance and phase transition of *Schistocerca gregaria* (Orthoptera: Acrididae). Egypt. Acad. J. biolog. Sci. 3(2), 133-147(2010).
- [33] Ghoneim K., Amer M., Al-Daly A., Mohammad A., Khadrawy F. and Mahmoud M., Disrupted survival, growth and development of desert locust *Schistocerca gregaria* (Forsk.) (Orthoptera: Acrididae) by extracts from toothpick weed *Ammi visnaga* Lamarck (Apiaceae). Int. J. Biosci. 5(1), 397-414(2014).
- [34] Bhardwaj A.K. and Ansari B.A., Effect of Nimbecidine and Neemazal on the developmental programming of cotton pest, *Earias vittella*. J.Entomol. Zool. Studies 3(1), 38-42(2015).
- [35] Truman J.W. and Riddiford L.M., Neuroendocrine control of ecdysis in silk moths. Sci. 167, 1624-1626(1970).
- [36] Truman J.W., Interaction between ecdysteriod, eclosion hormone and bursicon titres in *Manduca sexta*. Amr. Zool. 21, 655-661(1981).
- [37] Nicol C.M.Y. and Schmutterer H., Contact effects of seed oil from the neem tree *Azadirachta indica*, on nymphs of the gregarious phase of the desert locust, *Schistocerca gregaria* (Forsk.). J. Appl. Entomol. 111(2), 197-205(1991).
- [38] Osman M.Z., Effects of neem seed extract on growth and development of larvae of *Pieris brassicae* L (Lep.: Pieridae). J. App. Entomol. 115, 254-258(1993).
- [39] Ghoneim K.S., Mohamed H.A. and Bream A.S., Efficacy of the neem seed extract NeemAzal on the growth and development of the Egyptian cotton leafworm, *Spodoptera littoralis* Boisid (Lepidoptera: Noctuidae). J. Egypt. Ger. Soc. Zool. 33(E), 161-179(2000).
- [40] von Elling K., Borgemeister C., Setamou M. and Poehling H.M., Effect of Neemazal-T/S, a commercial neem product, on different developmental stages of the common greenhouse whitefly *Trialeurodes vaporariorum* Westwood (Hom., Aleyrodidae). J. App. Entomol. 126, 40-45(2002).
- [41] Athanassiour C.G., Kontodimas D.C., Kavallieratos N.G. and Veroniki M.A., Insecticidal effect of NeemAzal against three stored product beetle species on rye and oats. J.Econ. Entomol. 98, 1733-1738(2005).
- [42] Senthil-Nathan S., Kalaivani K., Chung K. and Murugan K., The toxicity and behavioural effects of neem limonoids on *Cnaphalocoris medinalis* (Guenee) the rice leafhopper. Chemosphere 62 (8), 1381-1387(2006).
- [43] Senthil Nathan S., Choi M.Y., Paik C.H., Seo H.Y., Kim J.D. and Kang S.M., The toxic effects of neem extract and azadirachtin on the brown planthopper, *Nilaparvata lugens* (Stal) (BPH) (Homoptera: Delphacidae). Chemosphere 67, 80-88(2007).

- [44] Siriwattananurongsee S., Sukontason K.L., Olson J.K., Chailapakul O. and Sukontason K., Efficacy of neem extract against the blowfly and housefly. *Parasitol. Res.* 103, 535-544(2008).
- [45] Tripathy A., Samanta L., Das S., Parida S.K., Marai N., Hazra R.K., Mallavdani U.V., Kar S.K. and Mahapatra N., The mosquitocidal activity of methanolic extracts of *Lantana camara* root and *Anacardium occidentale* leaf: role of glutathione S-transferase in insecticide resistance. *J. Med. Entomol.* 48(2), 291-295(2011).
- [46] Janakan R. and Ramakrishnan N., Mosquitocidal activity of *Barleria prionitis* Linn (Acanthaceae) and *Ageratum conyzoides* Linn (Asteraceae) against malarial vector mosquito *Anopheles stephensi* Liston (Diptera: Culicidae). *Int. J. Current Innov. Res.* 1(10), 45-50(2014).
- [47] Janakan, R. and Ramakrishnan, N., Mosquito Larvicidal and Adulticidal Properties of Botanical Extract *Barleria prionitis* Linn (Acanthaceae) Against *Aedes aegypti* Linn and *Culex quinquefasciatus* Say (Diptera: Culicidae). *Int. J. Current Innov. Res.* 1(11), 39-44(2014).
- [48] Naqvi S.N.H. and Parveen F., Toxicity and residual effect of *Nerium indicum* crude extract as compared with Coopex against adults *Tribolium castaneum* (Coleoptera: Tenebrionidae). *Pak. J. Entomol.*, Karachi 6, 35-44(1991).
- [49] Ghoneim K.S. and Al-Dali A.G., Survival and reproductive responses of *Muscina stabulans* Fallen (Muscidae: Diptera) to the neem seed extract, Margosan-O. *Proc. 2nd Int. Conf., Plant Prot. Res. Inst. Cairo, Egypt, 21-24 Dec. (2002).*
- [50] Amer M.S., Ghoneim K.S., Al-Dali A.G., Bream A.S. and Hamadah Kh. Sh., Assessment of the activity of Margosan-O and Jojoba against the house fly *Musca domestica* (Diptera: Muscidae). *Al-Azhar Bull. Sci.* 15(2), 09-24(2004).
- [51] Kabarou J.M. and Gichia L., Insecticidal activity of extracts derived from different parts of the mangrove tree *Rhizophora mucronata* (Rhizophoraceae) Lam. against three arthropods. *African J.Sci. Technol. (Science and Engineering Series)* 2(2), 44-49(2001).
- [52] Ghoneim K., Amer M., Al-Daly A., Mohammad A., Khadrawy F. and Mahmoud M., Effectiveness of *Punica granatum* Linn. (Lythraceae) extracts on the adult performance of desert locust *Schistocerca gregaria* (Forsk.) (Orthoptera: Acrididae). *Entomol. App. Sci. Letters*, 1(2): 9-19(2014).
- [53] Burits M. and Bucar F., Antioxidant activity of *Nigella sativa* essential oil. *Phytother. Res.* 14(5), 323-328(2000).
- [54] Al-Ghamdi M.S., The anti-inflammatory, analgesic and antipyretic activity of *Nigella sativa*. *J. Ethnopharmacol.* 76(1), 45-48(2001).
- [55] Ali B.H. and Blunden G., Pharmacological and toxicological properties of *Nigella sativa*. *Phytother. Res.* 17(4), 299-305(2003).
- [56] Ali M.A., Sayeed M.A. Alam M.S., Yeasmin M.S., Khan A.M. and Muhamad I.I., Characteristics of oils and nutrient contents of *Nigella sativa* Linn. and *Trigonella foenum-graecum* seeds. *Bull. Chem. Soc. Ethiop.* 26, 55-64(2012).
- [57] Schmutterer H., Baumgart M., Freisewikel D., Langenwald J. and Nicol C.M.Y., The effects of neem oil and other neem products on nymph and resulting adults of *Schistocerca gregaria*, *Nomadacris septemfasciata*, *Locusta migratoria migratorioides* and *Zonocerus variegates*. *J. App. Entomol.* 116(2), 178-186(1993).
- [58] El-Sokkary Z.F.A., Biological and physiological effects of some insect growth regulators and botanicals on the desert locust *Schistocerca gregaria* Forskal. *M.Sc. Thesis, Fac.Sci., Ain Shams Univ. Cairo, Egypt* (2003).
- [59] Nascimento I.R., Murata A.T., Bortoli S.A. and Lopes L.M., Insecticidal activity of chemical constituents from *Aristolochia pubescens* against *Aticarsia gemmatalis* larvae. *Pestic. Manage. Sci.* 60, 413-416(2004).
- [60] Cespedes C.L., Salazar J.R., Martinez M. and Aranda E., Insect growth regulatory effects of some extracts and sterols from *Myrtillocactus geometrizans* (Cactaceae) against *Spodoptera frugiperda* and *Tenebrio molitor*. *Phytochem.* 66, 2481-2493(2005).
- [61] Josephraj Kumar, A., Subrahmanyam, B. and Srinivasan, S., Plumbagin and azadirachtin deplete haemolymph ecdysteroid levels and alter the activity profiles of two lysosomal enzymes in the

- fat body of *Helicoverba armigera* (Lepidoptera: Noctuidae). Euro. J. Entomol. 96, 347-353(1999).
- [62] Uvarov B., Grasshoppers and Locusts, vol. 2. Centre for Overseas Pest Research, London, UK (1977).
- [63] Roessingh P., Simpson S.J. and James S., Analysis of phase-related changes in behavior of desert locust nymphs. Proc. R. Soc. B 252, 43-49(1993).
- [64] Tawfik A.I., Tanaka S., de Loof A., Schoofs L., Baggerman G., Waelkens E., Derua R., Milner Y., Yerushalmii Y. and Pener M.P., Identification of the gregarization-associated dark pigmentotropin in locusts through an albino mutant. Proc. Natl. Acad. Sci. USA 96, 7083-7087(1999).
- [65] Rogers S.M., Matheson T., Sasaki K., Kendrick K., Simpson S.J. and Burrows M., Substantial changes in central nervous system neurotransmitters and neuro-modulators accompany phase change in the locust. J. Exp. Biol. 207, 3603-3617(2004).
- [66] Pener M.P. and Simpson S.J., Locust phase polyphenism: an update. Adv. Insect Physiol. 36, 1-286(2009).
- [67] Gordon, S.D., Rogers, S. and Windmill, J., Hearing differences of gregarious and solitary locusts (*Schistocerca gregaria*), an example of epigenetic effects. Front. Behav. Neurosci. Conference: Tenth International Congress of Neuroethology, College Park. Maryland USA, USA, 5 Aug -10 Aug, 2012. (Abstracts) (2012).
- [68] Harano K.-I., Tanaka S., Watari Y. and Saito O., Phase-dependent locomotor activity in first-stadium nymphs of the desert locust, *Schistocerca gregaria*: Effects of parental and progeny rearing density. J. Insect Physiol. 58, 718-725(2012).
- [69] Pener M.P., Endocrine aspects of phase polymorphism in locusts. In: "Endocrinology of Insects". (Downer R.G.H. and Laufer H., eds.). New York, pp. 379-394(1983).
- [70] Rembold H. and Mwangi R.W., Compounds from *Melia volkensii* and their growth inhibitory effect on *Aedes aegypti* larvae. In: "Host regulated development mechanisms in vector arthropods" (Borovsky D., ed.). Spielman, A, pp: 3-8(1989).
- [71] Nasseh O., Wilps H., Rembold H. and Krall S., Biologically active compounds in *Melia volkensii*: Larval growth inhibitor and phase modulator against the desert locust *Schistocerca gregaria* (Forsk.) (Orth., Cyrtacanthacriniae). J. App. Entomol. 116(1), 1-11(1993).
- [72] Langewald J., Scherer R. and Schmutterer H., Repellent effects of different products of the neem tree on the red locust *Nomadacris septemfasciata* Serv. in maize fields in the southwestern parts of Madagascar. Anzeiger-fur- Schadlingskunde, -Pflanzenschutz, Umweltschutz. 68(3), 55-57(1995).
- [73] Schmutterer H. and Freres T., Influence of neem-seed oil on metamorphosis, colour and behaviour of the desert locust *Schistocerca gregaria* (Forsk.) and the African migratory locust *Locusta migratoria migratorioides* (R. & F.). Zeitschrift Fur Pflanzenkrankheiten Und Pflanzenschutz. 97(4), 431-438(1990).
- [74] Bakr R.F.A., Hussein M.A., Hamouda L.S., Hassan H.A. and Elsokary Z.F., Effect of some insecticidal agents on some biological aspects and protein patterns of desert locust, *Schistocerca gregaria* (Forsk.). Egypt. Acad. Soc. Environ. Develop. 9(2), 29-42(2008).
- [75] Richard D.S., Jones J.M., Barbarito M.R., Cerula S., Detweiler J.P., Fisher S.J., Brannigan D.M. and Scheswohl D.M., Vitellogenesis in diapausing and mutant *Drosophila melanogaster*: further evidence for the relative roles of ecdysteroids and juvenile hormones. J. Insect Physiol. 47, 905-913(2001).
- [76] Uvarov B.P., Grasshoppers and locusts. A Handbook of General Acridology, I. The Univ. Press, Cambridge, 481 pp. (1966).
- [77] Nolte, D.J., A pheromone for melanization of locusts. Nature 16, 660-661(1963).
- [78] Gillett S.D. and Phillips M.L., Faeces as a source of a locust gregarization stimulus. Effects on social aggregation and on cuticular colour of nymphs of the desert locust, *Schistocerca gregaria* (Forsk.) Acrida, 6: 279-286(1977).
- [79] Jacobson M., Botanical pesticides, past, present and future. In: "Insecticides of plant origin" (Arnason J.T., ed.). Proc. Am. Chemical Soc. Washington, DC, 1-10(1989).

- [80] Hamadah Kh. Sh., Physiological and Biochemical Effects of IGRs and plant extracts on the house fly *Musca domestica*. M.Sc. Thesis, Fac. Sci., Al-Azhar Univ., Cairo, Egypt (2003).
- [81] Basiouny A.L.I., Reproductive responsiveness of *Schistocerca gregaria* (Orthoptera: Acrididae) to different extracts from the wild plant *Fagonia bruguieri*. J.Biol. Pharm. Sci. 6(1), 178-193(2008).
- [82] Abou El-Ela R.G., Helmy N.M., El Monairy O.M. and Salah H., Biological activity of an extract from *Hyoscyamus muticus* on *Musca domestica*. (Diptera, Muscidae). Bull. Entomol. Soc. Egypt, (Econ. Ser.) 22(17), 27-35(1995).
- [83] Muse W.A., Lajide L. and Adedire C.O., Effects of some Nigerian plants on survival, oviposition, and emergence of adult blowfly, *Chrysoma chloropyga* (Wied.) (Diptera: Calliphoridae). J. Asia-Pacific Entomol. 6(1), 69-75(2003).
- [84] Steffens R.J. and Schmutterer H., The effect of a crude methanolic neem (*Azadirachta indica*) seed kernel extract on metamorphosis and quality of adults of the Mediterranean fruit fly, *Ceratitis capitata* Wied. (Diptera: Tephritidae). Z Angew Entomol. 94, 98-103(1982).
- [85] Gujar G.T. and Mehrotra K.N., Inhibition of growth and development of the tobacco caterpillar, *Spodoptera litura* Fabr. Due to Azadirachtin and other neem products. Indian J. Entomol. 45, 431-435(1983).
- [86] Gujar G.T. and Mehrotra K.N., Juvenilizing effect of Azadirachtin on a noctuid moth, *Spodoptera litura* Fabr. Indian J. Exp. Biol. 21, 292-293(1983).
- [87] Mehrotra K.N. and Gujar G.T., Neem as insect growth inhibitor. Natn. Seminar on "Neem in Agriculture", IARI, Neem Newsl. 1, 6(1984).
- [88] Di Ilio V., Cristofaro M., Marchini D., Nobili P. and Dallai R., Effects of a neem compound on the fecundity and longevity of *Ceratitis capitata* (Diptera: Tephritidae). J. Econ. Entomol. 92, 76-82(1999).
- [89] Schmidt G.H., Ahmed A.A. and Breuer M., Effect of *Melia azedarach* extract on larval development and reproduction parameters of *Spodoptera littoralis* (Boisd) and *Agrotis ipsilon* (Hufn.) (Lep., Noctuidae). Anzeiger-Fur-Schadlingskunde, -Pflanzenschutz, Umweltschutz. 70(1), 04-12(1997).
- [90] Hassan H.A., Biological and biochemical studies on the effects of some botanical extracts on cotton leafwom *Spodoptera littoralis* (Boisd.) (Lepidoptera: Noctuidae). M.Sc. Thesis, Fac. Sci., Ain Shams Univ., Cairo, Egypt (2002).
- [91] Hamadah Kh. Sh., Some developmental, haematological and enzymatic effects of certain plant extracts on the desert locust *Schistocerca gregaria* (Orthoptera: Acrididae). Ph.D. Thesis, Al-Azhar Univ., Cairo, Egypt (2009).
- [92] Clancy D.J., Gems D., Harshman L.G., Oldham S., Stocker H., Hafen E., Leivers S.J. and Partridge L., Extension of life-span by loss of CHICO, a *Drosophila* insulin receptor substrate protein. Sci. 292, 104-106(2001).
- [93] Simon A.F., Shih C., Mack A. and Benzer S., Steroid control of longevity in *Drosophila melanogaster*. Sci. 299, 1407-1410(2003).
- [94] Broughton S.J., Piper M.D., Ikeya T., Bass T.M., Jacobson J., Driege Y., Martinez P., Hafen E., Withers D.J., Leivers S.J. and Partridge L., Longer lifespan, altered metabolism, and stress resistance in *Drosophila* from ablation of cells making insulin-like ligands. Proc. Natl. Acad. Sci. U.S.A. 102, 3105-3110(2005).
- [95] Yamamoto R., Bai H., Dolezal A.G., Amdam G. and Tatar M., Juvenile hormone regulation of *Drosophila* aging. BMC Biology 11, 85(2013).

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