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Scintigraphic Changes in Experimental Diabetes

Bioavailability of pertechnetate in diabetes Irami Araújo-Filho^{a*}, Amália Cinthia Meneses Rêgo^a

^aFull Professor of the Graduate Program in Biotechnology, Potiguar University,
Brazil - Laureate International Universities (UnP), Natal - RN, Brazil;
Ph.D in Health Sciences from the Federal University of Rio Grande do Norte/NATAL/BRAZIL.

^{a*}Associate Professor in Surgery Department of the Federal University of the Rio Grande do Norte

irami.filho@uol.com.br

Abstract: In nuclear medicine practice, physicians set norms for morphological or physiological function for each organ by diagnosing a large number of patients. For every procedure, there is diagnostic data for a range of normal variations familiar to physicians. This study aimed to clarify if diabetes induced in rats changes the biodistribution of the radiopharmaceutical sodium pertechnetate (Na TcO4) and, consequently, the accuracy of the scintigraphic exams. We used 14 male Wistar rats, randomly allocated in 2 group: the diabetic group (D) rats (n=7) were submitted to the induction of diabetes with streptozotocin, and 7 non diabetic rats were used as controls (C). After 7 days, in the 14 animals it was administered 0,1mL of Na TcO4 (0.66 MBq) through orbital plexus and, after 30 minutes, the radiopharmaceutical sodium pertechnetate was evaluated, being compared the uptske of this in the several studied organs of diabetic animals and controls. The biodistribution of Na TcO4 was significantly higher in the liver and smaller in the bladder, thyroid and stomach of diabetic mice when compared to the control group rats. Diabetes induced in rats alters the biodistribution of Na TcO4, and this finding could have clinical implications on scintigraphic exams.

Keywords: Bioavailability. Tc 99m Pertechnetate. Diabetes. Streptozotocin. Scintigraphy. Rats.

1. Introduction

The radionuclides are employed in many fields of knowledge. In health sciences they contribute to the improvement of diagnosis and treatment, enabling advances in clinical and experimental research. Most diagnostic tests like scintigraphy, reveals the uptake of radioisotopes in several organs and tissues [1-4]. The most used radioisotopes are gamma radiation emitting, as a source of radiation or tracers [1].

Since the 60s, the 99m Technetium (99m Tc) is used in the biomedical area because it is easily obtained from molybdenum/technetium (99 Mo/ 99 mTc) generators, it has short half-life (6h), low emission energy, it is easy to label red blood cells, cellular structures or molecular, it has low cost and low environmental impact [5-8]. These features make 99m Tc the most widely used radioisotope, administered in the form of sodium pertechnetate (Na 99m TcO4) or attached to other molecules. Their biodistribution is evaluated by scintigraphy or other systems to detect radioactivity [6,7].

The scintigraphic image reflects morphology and organ function. Thus, different radioactive compounds can be used to study the physiology of organs and tissues. Therefore, a diseased tissue maycontain higher or lower uptake of radioactivity, depending on its disfunction [8]. Scintigraphic examinations are used in the diagnosis of tumors, metastasis, gastric emptying, esophageal motility, thyroid nodules, bleeding, and in the monitoring of systemic diseases and their consequences, among them, diabetes [9-15].

The metabolic imbalance associated with diabetes causes secondary disorders in multiple organ systems, which can cause alterations in biodistribution of radioisotopes. Changes in scintigraphic examination results can generate false positive images, repetition of exams and increased patient exposure to radiation. Some studies advocate the use of streptozotocin to induce experimental diabetes, because it is a cytotoxic chemical agent, specific for pancreatic beta cells, causing severe

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primary insulin insuficiency and high glucose levels, followed by the establishment of diabetes in the subsequent 24 hours. This model, although with a high mortality rate is relatively easy, destroying the endocrine cells with preservation of exocrine function of pancreas [16-18].

Considering such principles, the objective of this study was to evaluate the bio distribution of sodium pertechnetate (Na^{99m}TcO4) in different organs of rats with diabetes.

2. MATERIALS AND METHODS

We used 14 three months old Wistar rats, weighing $265g \pm 31g$, provided by the vivarium of the Health Sciences Center, Federal University of Rio Grande do Norte, Brazil. All animals were weighed, placed in individual cages with water and food (Purina[®] Labina) *ad libitum* and acclimated in the laboratory for 7 days. They were kept under controlled temperature (21 degrees), humidity (60-70%), lighting (12/12 h) light / dark cycle and handled in accordance with the Ethical Code for Animal Experimentation (Council for International Organization of Medical Sciences) and the rules of the Brazilian College of Animal Experimentation. They were distributed randomly into two groups: diabetes (D, n = 7) and control group (C, n = 7). The animals in group D were subjected to induction of diabetes by injecting streptozotocin at the dose 80 mg/kg, intraperitoneally (IP). Seven days after the injection of streptozotocin, glucose was measured by collecting blood from the dorsal vein of the tail, using the equipment Accu-Chek Advantage, Roche Diagnostics[®], Mannheim, Germany (2003). We considered diabetic the animals with fasting glucose above 200 mg/dL. The C animals were not subjected to any procedure, only observed.

On day 7, all animals were anesthetized with thiopental (20mg/kg-IP) and Ketamine (20mg/kg-IM), administered 0,1mL Na 99m TcO4 intravenously into the orbital plexus, and radioactivity dose was 0.68MBq. After 30 minutes, the animals were killed with a lethal dose of anesthetic thiopental (100mg/kg), intracardiac, and samples were harvested from pancreas, brain, thyroid, lung, heart, stomach, liver, kidney, bladder and right femur. The samples were washed in 0.9% NaCl, weighed on a digital precision balance (Bel-Mark-160-II-Italy $^{\Box}$) and taken for detection of radioactivity through the 1470 Auto Gamma counter, WizardTM-Perkin-Elmer $^{\textcircled{\$}}$ (Finland) with automatic correction of decay. The percentage of radioactivity per gram ($^{\textcircled{\$}}$ ATI/g) of each organ was calculated by dividing the activity per gram of tissue by the total radioactivity administered to each animal. The experiment was completely randomized with statistical analysis by Student t test. The level of significance for the test was 5% (95% CI), ie, p<0.05 was considered statistically significant.

3. RESULTS & DISCUSSION

Table 1 shows the results of %ATI/g in group D and C. In group D rats we observed a significant increase of the biodistribution of Na99mTcO4 in the liver and reduction in the uptake of thyroid, bladder and stomach when compared with group C rats. (p <0.05). The other organs did not show difference in %ATI/g, comparing with controls.

Table 1 Diadiatribution of Na 99n	ⁿ TcO4 in organs and respective groups
Table1. Bioaistribution of Na	1 cO4 in organs and respective groups

Organ	Diabetes	Control	<i>p</i> -valor
Brain	0.02 ± 0.008	0.01 ± 0.003	0.093109
Thyroid	0.46 ± 0.154	3.89 ± 1.447	0.00075*
Lung	0.41 ± 0.157	0.39 ± 0.141	0.803036
Heart	0.23 ± 0.067	0.29 ± 0.061	0.223256
Stomach	0.43 ± 0.373	3.33 ± 1.593	0.004118*
Liver	2.97 ± 1.745	0.34 ± 0.050	0.009782*
Kidney	0.27 ± 0.161	0.42 ± 0.074	0.106062
Bladder	0.19 ± 0.055	0.34 ± 0.094	0.015865*
Femur	0.15 ± 0.058	0.14 ± 0.032	0.709645
Pancreas	0.15 ± 0.047	0.13 ± 0.046	0.518932

Mean \pm *SD.* **Significant diference* p<0.05.

1- p-valor of Student t test.

Research shows the interference of anesthetic drugs, chemotherapy and herbals on the biodistribution of sodium pertechnetate (Na^{99m}TcO4). Recently, studies from our laboratory demonstrated changes in the uptake of radiopharmaceuticals in the postoperative of major surgeries involving experimental models of short bowel syndrome and bariatric surgery[19-23].

Diabetes mellitus and associated metabolic disturbances interfere with homeostasis, which may have caused changes in the biodistribution of pertechnetate on this study. We detected an increase in radioisotope uptake in the liver and a reduction in the stomach, bladder and thyroid gland of diabetic rats compared to controls. Bertin et al. argued that the control of gastric emptying involves neurological, hormonal and metabolic factors and that the role of diabetes on gastric motility is little known [24]. However El-Shaldy et al. attributed the delay in gastric emptying to the inhibition of the hormone motilin by hyperglycemia [25]. Theoretically, it was expected to obtain a higher uptake of gastric pertechnetate in rats of the experimental group, given the prior knowledge of several other trials on diabetic gastroparesis associated with autonomic neuropathy. However, we observed the opposite. This may affect the results of scintigraphy for the study of gastric emptying in diabetic patients commonly performed during the search of dispeptic-symptoms [14, 26-31].

Another autonomic dysfunction in diabetes is the neurogenic bladder. Many researchers adopt as diagnostic of this pathology an increased tracer retention in the bladder during the performance of radionuclide cystography. The bladder dysfunction would result from neurological, muscular and urothelial injuries, uriginated on hyperglycemia [32,33].

We suppose that the low bladder uptake of sodium pertechnetate observed in this study was due to a marked polyuria in the experimental group rats, which may have contributed to greater intravesical radioactive energy dissipation. In relation to thyroid disorders associated with diabetes, it is a phenomenon widely reported in the current literature. It is estimated that diabetes is associated with hypothyroidism, Hashimoto's thyroiditis and nontoxic goiter in about 20-56% of cases involving mainly patients with type II diabetes [34-36]. It is therefore of great importance for thyroid evaluations. In this sense, thyroid scintigraphy is used in complementing clinical data and ultrasound.

The use ^{99m}Tc displays advantages over iodine ¹³¹, because it labels the gland, allows its morphofunctional evaluation, with less patient exposure to radiation without interfering with the glandular function [36].

There was a lower thyroid uptake of sodium pertechnetate (Na ^{99m}TcO4) in our diabetic animals due to a probable subclinical hypothyroidism, as evidenced by Akbar et al, studying thyroid dysfunction in patients with type II diabetes [37]. Passos et al. stated that a reduction in the synthesis of thyroglobulin precursor results in a lower uptake of sodium pertechnetate by the gland, since there is not enough protein to be marked. Thus, we alert to possible changes in thyroid scintigraphy in diabetic patients [38].

A final analysis concerns the greater hepatic uptake of pertechnetate in the rats of the experimental group. Clinical trials point to the diabetes in the pathogenesis of liver diseases, with emphasis on non-alcoholic steatohepatitis, cirrhosis and hepatocellular carcinoma, including transplanted patient [39-45] The liver participates directly in the glucose metabolism, raising or lowering the glucose uptake as vary their organic levels. It is postulated that a normalization of blood glucose, would result in regression of these pathological processes in inicial stage [46]. In this study, a likely steatohepatitis could have caused an increased uptake of sodium pertechnetate in the liver of diabetic rats, because once present in the circulation, the radiopharmaceuticals label not only erythrocytes but also leukocytes, showing a higher bioavailability in inflammed sites [47,48]. This phenomenon could alter the results of liver scintigraphy when searching for primary tumors, liver cirrhosis and functional assessments in detecting metastases.

4. CONCLUSION

Diabetes altered the biodistribution of sodium pertechnetate (Na^{99m}TcO4) in the liver, stomach, thyroid and urinary bladder of rats, emphasizing the need for a careful analysis of the results, if scintigraphic examinations are to be performed in patients with this disease.

REFERENCES

- [1] Sampsom CB. Adverse reactions and drug interactions with radiopharmaceuticals. Drug Safety. 1993;8:280-94.
- [2] Gutfilen B, Pellini MP, Roure-Neder J, Amarante Júnior JL, Evangelista MG, Fernandes SR, et al. 99mTc labeling white blood cells with a simple technique: clinical application. Ann Nucl Med. 1994;8:85-9.
- [3] Moreno SRF, Mattos MF, Rocha EK, Khan MM, San Gil Rãs, Conceição RC, et al. Thalidomide: Labelling and suggestion of the chemical binding with techenetion-99m. Chem Nucl Med. 1999;65:641-6.
- [4] Gutfilen B, Ribeiro BLAR, Mattos MF, Ribeiro CR, Bernardo-Filho M. Labeling of thymidine technetion-99: suggestion of a chemical model. Braz Arch Biol Tecnol.1996;39:69-74.
- [5] Saha GB. Fundamentals of nuclear Pharmacy. 4ed. New York:Springer-Verlag; 1997.
- [6] Hladik WB, Saha GB. Study T. Essentials of Nuclear Medine Science. Baltimore: Williams; 1987: p3-50.
- [7] Braga AC, Oliveira MB, Feliciano GD, Reiniger IW, Oliveira JF, Silva CR et al. The effect of drugs on the labeling of blood elements with technetium-99m. Curr Pharm Des. 2000;6:1179-91.
- [8] Saha GB. Fundamentals of nuclear pharmacy. New York:Springer-Verlag; 2004.
- [9] Buchmann I, Riedmüller K, Hoffner S, Mack U, Aulmann S, Haberkorn U. Comparison of 99mtechnetium-pertechnetate and ¹²³iodide SPECT with FDG-PET in patients suspicious for breast cancer. Cancer Biother Radiopharm. 2007;22:779-89.
- [10] Hansen AA, Rosenberg RJ. Gastrointestinal stromal tumor detected on Tc- 99m red blood cell scintigraphy. Clin Nucl Med. 2007;32:221-3.
- [11] Shie P, Cardarelli R, Brandon D, Erdman W, Abdulrahim N. Meta-analysis: comparison of F-18 Fluorodeoxyglucose-positron emission tomography and bone scintigraphy in the detection of bone metastases in patients with breast cancer. Clin Nucl Med. 2008;33:97-101.
- [12] Mariani G, Boni G, Barreca M, Bellini M, Fattori B, AlSharif A, et al. Radionuclide gastroesophageal motor studies. J Nucl Med. 2004;45:1004-28.
- [13] Stathaki MI, Karkavitsas NS. Nuclear Medicine in the diagnosis of lower gastrointestinal bleeding. Hell J Nucl Med. 2007;10:197-204.
- [14] Faraj J, Melander O, Sundkvist G, Olsson R, Thorsson O, Ekberg O, et al. O esophageal dysmotility, delayed gastric emptying and gastrointestinal symptoms in patients with diabetes mellitus. Diabet Med. 2007;24:1235-9.
- [15] Ohlsson B, Melander O, Thorsson O, Olsson R, Ekberg O, Sundkvist G. O esophageal dysmotility, delayed gastric emptying and autonomic neuropathy correlate to disturbed glucose homeostasis. Diabetologia. 2006;49:2010-4.
- [16] Delfino VDA, Figueiredo JF, Matsuo T, Fávero ME, Matni AM, Mocelin AJ. *Diabetes mellitus* induzido por estreptozotocina: comparação em longo prazo entre duas vias de administração. J Bras Nefrol. 2002;24:31-6.
- [17] Taniguchi H, Muroi R, Kobayashi-Hattori K, Uda Y, Oishi Y, Takita T. Differing effects of water-soluble and fat-soluble extracts from Japanese radish (Raphanus sativus) sprouts on carbohydrate and lipid metabolism in normal and streptozotocin-induced diabetic rats. J Nutr Sci Vitaminol. 2007;53:261-6.
- [18] Ozkaya YG, Ağar A, Hacioğlu G, Yargiçoğlu P. Exercise improves visual deficits tested by visual evoked potentials in streptozotocin-induced diabetic rats. Tohoku J Exp Med. 2007;213:313-21.
- [19] Simões SB, Machado-Silva JR, Gutfilen B, Presgrave OA, Oliveira MB, Bernardo Filho M. Biodistribution study of the anaesthetic sodium phenobarbital [20] labelled with technetium-99m in Swiss mice infected with Schistosoma mansoni Sambon, 1907. Mem Inst Oswaldo Cruz. 1997;92:677-81.
- [20] Santos JS, de-Paula EF, Correa TG, de-Freitas LC, da-Fonseca LM, Gutfilen B, et al. Effect of cyclophosphamide on the binding of 99mTcO-4 and 99mTc-MDP to blood cells and plasma proteins. Braz J Med Biol Res. 1995;28:131-5.

- [21] Oliveira JF, Avila AS, Braga AC, de Oliveira MB, Boasquevisque EM, Jales RL, et al. Effect of extract of medicinal plants on the labeling of blood elements with Technetium-99m and on the morphology of red blood cells: I--a study with *Paullinia cupana*. Fitoterapia. 2002;73:305-12.
- [22] Chacon DA, Araújo-Filho I, Villarim-Neto A, Rêgo AC, Azevedo IM, Bernardo-Filho M et al. Biodistribution of the radiophamarceutical sodium pertechnetate (Na99mTcO4) after massive small bowel resection in rats. Acta Cir Bras. 2007;22:430-5.
- [23] Araújo-Filho I, Rêgo ACM, Brandão-Neto J, Villarim-Neto A, Egito EST, Azevedo IM et al . Biodistribution of the radiopharmaceutical sodium pertechnetate after biliopancreatic bypass with a duodenal switch. Braz Arch Biol Technol. 2007;50:189-97.
- [24] Bertin E, Schneider N, Abdelli N, Wampach H, Cadiot G, Lobo Guerrero A, et al. Gastric emptying is accelerated in obese type 2 diabetic patients without autonomic neuropathy. Diabetes Metab. 2001;27:357-64.
- [25] El-Salhy M, Spångéus A. Gastric emptying in animal models of human diabetes: correlation to blood glucoselevel and gut neuroendocrine peptide content. Ups J Med Sci. 2002;107:89-99.
- [26] Malmud LS, Fisher RS, Knight LC, Rock E. Scintigraphic evaluation of gastric emptying. Semin Nucl Med.1982;12:116-25.
- [27] Cesarini PR, Ferreira SRG, Dib SA. Gastroparesia diabética. Rev Assoc Med Bras. 1997;43:163-8.
- [28] Jones KL, Horowitz M, Wishart MJ, Maddox AF, Harding PE, Chatterton BE. Relationships between gastric emptying, intragastric meal distribution and blood glucose concentrations in diabetes mellitus. J Nucl Med. 1995;36:2220-8.
- [29] Nowak TV, Johnson CP, Kalbfleisch JH, Roza AM, Wood CM, Weisbruch JP, et al. Highly variable gastric emptying in patients with insulin dependent diabetes mellitus. Gut. 1995;37:23-9.
- [30] Qi HB, Luo JY, Zhu YL, Wang XQ. Gastric myoelectrical activity and gastric emptying in diabetic patients with dyspeptic symptoms. World J Gastroenterol.2002;8:180-2.
- [31] Yoshimura N, Chancellor MB, Andersson KE, Christ GJ. Recent advances in understanding the biology of diabetes-associated bladder complications and novel therapy. BJU Int. 2005;95:733-8.
- [32] Nagabhushan N, Syed R, Hoh IM, Syed I, Ell PJ, Shah PJ, et al. 99mTechnetium-mercaptoacetyltriglycine scintigraphy with full bladder in patients with severe bladder dysfunction. J Urol. 2006;176:1481-6.
- [33] Pimenta WP, Mazeto GMFS, Callegaro CF, Shibata SA, Marins LV, Yamashita S, et al. Associação de tireopatias em uma população de pacientes com diabetes. Arq Bras Endocrinol Metab. 2005;49:234-40.
- [34] Umpierrez GE, Latif KA, Murphy MB, Lambeth HC, Stentz F, Bush Al. Thyroid dysfunction in patients with type 1 diabetes: a longitudinal study. Diabetes Care. 2003;26:1181-5.
- [35] Chubb SA, Davis WA, Davis TM. Interactions among thyroid function, insulin sensitivity, and serum lipid concentrations: the Fremantle diabetes study. J Clin Endocrinol Metab. 2005;90:5317-20.
- [36] Ramos CD, Zantut Wittmann DE, Etchebehere EC, Tambascia MA, Silva CA, Camargo EE. Thyroid uptake and scintigraphy using 99mTc pertechnetate: standardization in normal individuals. Sao Paulo Med J. 2002;120:45-8.
- [37] Akbar DH, Ahmed MM, Al-Mughales J. Thyroid dysfunction and thyroid autoimmunity in Saudi type 2 diabetics. Acta Diabetol. 2006;43:14-8.
- [38] Passos MC, Ramos CF, Bernardo-Filho M, de Mattos DM, Moura EG. The effect of protein or energy restriction on the biodistribution of Na99TcmO4 in Wistar rats. Nucl Med Commun. 2000;21:1059-62.
- [39] Hickman IJ, Macdonald GA. Impact of diabetes on the severity of liver disease. Am J Med. 2007;120:829-34.
- [40] Bell DS, Allbright E. The multifaceted associations of hepatobiliary disease and diabetes. Endocr Pract. 2007;13:300-12.

- [41] Komura T, Mizukoshi E, Kita Y, Sakurai M, Takata Y, Arai K. Impact of diabetes on recurrence of hepatocellular carcinoma after surgical treatment in patients with viral hepatitis. Am J Gastroenterol. 2007;102:1939-46.
- [42] Moscatiello S, Manini R, Marchesini G. Diabetes and liver disease: an ominous association. Nutr Metab Cardiovasc Dis. 2007;17:63-70.
- [43] Targher G, Bertolini L, Padovani R, Rodella S, Tessari R, Zenari L. Prevalence of nonalcoholic fatty liver disease and its association with cardiovascular disease among type 2 diabetic patients. Diabetes Care. 2007;30:1212-8.
- [44] Toledo FG, Sniderman AD, Kelley DE. Influence of hepatic steatosis (fatty liver) on severity and composition of dyslipidemia in type 2 diabetes. Diabetes Care. 2006;29:1845-50.
- [45] Iozzo P, Hallsten K, Oikonen V, Virtanen KA, Kemppainen J, Solin O. Insulin- mediated hepatic glucose uptake is impaired in type 2 diabetes: evidence for a relationship with glycemic control. J Clin Endocrinol Metab. 2003;88:2055-60.
- [46] Papathanasiou ND, Rondogianni PE, Pianou NK, Karampina PA, Vlontzou EA, Datseris IE. 99mTc-depreotide in the evaluation of bone infection and inflammation. Nucl Med Commun. 2008;29:239-46.
- [47] Joseph B, Bhargava KK, Tronco GG, Palestro CJ, Gupta S. Systemic and local release of inflammatory cytokines regulates hepatobiliary excretion of 99mTc- mebrofenin. Nucl Med Commun. 2008;29:336-44.
- [48] Carmo VA, Ferrari CS, Reis EC, Ramaldes GA, Pereira MA, De Oliveira MC. Biodistribution study and identification of inflammation sites using 99mTc-labelled stealth pH-sensitive liposomes. Nucl Med Commun. 2008;29:33-8.