

## **Dupuytren and Collagenase: Are they really as Simple as they Seem?**

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### **1. EDITORIAL**

In the treatment of Dupuytren's disease (DD), the use of Collagenase Clostridium Histolyticum (CCH) represents a new therapeutic alternative for this condition which currently has no definitive cure. Since the publication of the study by Hurst et al. [1] in which the immediate clinical results were very positive, several studies have validated the efficacy of CCH using different drug administration protocols with the same safety values [2-5]. Clinical results appear promising, with recurrence rates similar to those after surgery [6]. The cost-effectiveness ratio is favorable, compared to fasciectomy [7], and outpatient treatment improves the quality of healthcare for both orthopedic surgeons and for patients [8]. Functional recovery frequently takes place immediately, and evolution is generally more favorable.

However, some authors [9] find the rate of adverse events associated with CCH alarming. Although adverse events affect more than 85% of patients, most complications are minor, transient and mild [10]. Major and severe complications occur much less frequently than with other types of treatment, such as fasciectomy or dermofasciectomy.

There is little available material published on the mechanism of action and the adverse events associated with CCH. The CCH administered for DC consists of two isoforms: AUX I and AUX II, obtained from the purification of Clostridium Histolyticum toxins. CCH anchors in fibrillar collagen, especially types I and III, causing degradation and digestion. This leads to a chemical digestion of the DC cord [11]. Undoubtedly, the adverse events reported are related to treatment administration and the mechanism of action. But, what exactly is the mechanism of action? In a short communication, De Carlo [12] proposes that the inflammatory effects are a consequence of CCH administration. Likewise, studies performed in the 80s using Nucleolisyn® [13] with the use of CCH for the treatment of herniated disks and Peyronie's disease [14] also confirmed this process through an increase in vascular permeability and a healing response of the wound with inflammatory phenomena. The appearance of CCH adverse events is based on the facts that: A) CCH is a protein with bacterial and exogenous origin that activates the immunological mechanisms in the organism; and B) the degradation of collagen causes a response similar to the healing of any other wound. The latter process is the basis for much current investigation: collagen degradation activating the mediators for complete digestion [15] (endogenous metalloproteinases (MMP)). In turn, these MMP are regulated by certain inhibitors ( $\alpha$ 2-macroglobulin [16] and TIMPs [15]). This entire ensemble relates to molecules which, since they act on the extracellular matrix, may be considered paracrine factors (interleukins [17], IGF2 [18], etc.) under these circumstances. There is also interaction with molecules related to the cell surface (MT-MMPs or matrix metalloproteins) or the degradation of the rest of the extracellular matrix (ADAMTs acting as aggrecanases, for example [19]). The interrelations among these processes are very complex and variable, depending on the particular phase of Dupuytren's disease (cellular or nodular phases vs. acellular or fibrotic phases). Proof thereof is the fact that the research methods here under discussion have been applied to studies on the physiopathology Dupuytren's disease [20-23], the healing process of wounds [24-26], pathologic fibrosis [27-29], or cancer [30,31]. It should be noted that the processes involved are active both physiologically and pathologically. Indeed, the current situation regarding DC seems a bit chaotic. Apart from these areas of research into the processes within the cell [32] and those mentioned above that occur inside the extracellular matrix, there are other very different lines of investigation awaiting study: the

relationship of paracrine factors with regard to myofibroblasts, the relationship between myofibroblasts and the extracellular matrix [33], self-regulation of the extracellular matrix, distant factors or relationships such as the connection between DC and adhesive capsulitis [34], and others. Unifying all the various research lines is a complicated process, since professionals approaching the subject from the standpoint of so many different specialties frequently have little contact with each other (clinicians, pathologists, biochemists, orthopedic surgeons,...), and assessing results in clinical terms is difficult and leads to enormous uncertainties of opinion within the community of orthopedic surgeons. For example, there is no universal agreement as to the definition of DC recurrence [35]. Recent advances, such as the possibility of sustaining pathological DC tissue in live rats [36] may help to establish models for carrying out more uniform research. These multiple investigative lines are encouraging, and the advances are promising.

To conclude, much more research into the pharmacological aspects of CCH needs to be carried out, both regarding its positive side (the possibility of reapplication in patients after satisfactory initial treatment) and its negative side (analysis of adverse events and options to reduce them). In this regard the knowledge of the mechanism of action of the CCH is essential to development and approach to clinical trials. Knowing these mechanisms could help determine which developments after CCH administration are not complications, but rather processes intrinsic to the administration of CCH, as it has been similarly established for the surgical wounds in fasciectomy or for fat removal through liposuction in percutaneous aponeurotomy and lipofilling (PALF) [37].

#### REFERENCES

- [1] Hurst LC, Badalamente MA, Hentz VR, Hotchkiss RN, Kaplan FTD, Meals RA, et al. Injectable Collagenase Clostridium Histolyticum for Dupuytren's Contracture. *N Engl J Med* 361:968–79 (2009). doi:10.1056/NEJMoa0810866.
- [2] Atroshi I, Nordenskjöld J, Lauritzson A, Ahlgren E, Waldau J, Waldén M. Collagenase treatment of Dupuytren's contracture using a modified injection method. *Acta Orthop* 86:310–5 (2015). doi:10.3109/17453674.2015.1019782.
- [3] Coleman S, Gilpin D, Kaplan FTD, Houston A, Kaufman GJ, Cohen BM, et al. Efficacy and Safety of Concurrent Collagenase Clostridium Histolyticum Injections for Multiple Dupuytren Contracture. *J Hand Surg* 39:57–64 (2014). doi:10.1016/j.jhssa.2013.10.002.
- [4] Coleman S, Gilpin D, Tursi J, Kaufman G, Jones N, Cohen B. Multiple concurrent collagenase clostridium histolyticum injections to dupuytren's cords: an exploratory study. *BMC Musc Dis* 13:61 (2012). doi:10.1186/1471-2474-13-61.
- [5] Gaston RG, Larsen SE, Pess GM, Coleman S, Dean B, Cohen BM, et al. The Efficacy and Safety of Concurrent Collagenase Clostridium Histolyticum Injections for 2 Dupuytren Contracture in the Same Hand: a Prospective, Multicenter Study. *J Hand Surg* 40:1963–71 (2015). doi:10.1016/j.jhssa.2015.06.099.
- [6] Peimer CA, Blazar P, Coleman S, Kaplan FTD, Smith T, Lindau T. Dupuytren Contracture Recurrence Following Treatment With Collagenase Clostridium Histolyticum (CORDLESS [Collagenase Option for Reduction of Dupuytren Long-Term Evaluation of Safety Study]): 5-Year Data. *J Hand Surg* 40:1597–605 (2015). doi:10.1016/j.jhssa.2015.04.036.
- [7] Chen NC, Shauver MJ, Chung KC. Cost-Effectiveness of Open Partial Fasciectomy, Needle Aponeurotomy, and Collagenase Injection for Dupuytren Contracture. *J Hand Surg* 36:1826–34.e32 (2011). doi:10.1016/j.jhssa.2011.08.004.
- [8] Sanjuan-Cerveró R, Franco-Ferrando N, Poquet-Jornet J. Use of resources and costs associated with the treatment of Dupuytren's contracture at an orthopedics and traumatology surgery department in Denia (Spain): collagenase clostridium histolyticum versus subtotal fasciectomy. *BMC Musc Dis* 14:293 (2013). doi:10.1186/1471-2474-14-293.
- [9] Warwick DJ, Graham D, Worsley P. New insights into the immediate outcome of collagenase injections for Dupuytren's contracture. *J Hand Surg Eur Vol.* 25 (2015) [epub ahead of print] doi:10.1177/1753193415600670.
- [10] Peimer CA, Wilbrand S, Gerber RA, Chapman D, Szczypa PP. Safety and tolerability of collagenase Clostridium histolyticum and fasciectomy for Dupuytren's contracture. *J Hand Surg Eur Vol* 2014:1753193414528843. doi:10.1177/1753193414528843.

- [11] Crivello KM, Potter HG, Moon ES, Rancy SK, Wolfe SW. Does collagenase injection disrupt or digest the Dupuytren's cord: a magnetic resonance imaging study. *J Hand Surg Eur Vol* [epub ahead of print] 2016. doi:10.1177/1753193415626113.
- [12] Del Carlo M, Cole AA, Hart SGE, Levine LA. comparative analysis of collagen degradation in peyronie's disease plaque and dupuytren's contracture cord tissues injected with mixed collagenase subtypes. *J Urol* 181:279 (2009). doi:10.1016/S0022-5347(09)60794-1.
- [13] Bromley JW, Hirst JW, Osman M, Steinlauf P, Gennace RE, Stern H. Collagenase: an experimental study of intervertebral disc dissolution. *Spine* 5:126–32 (1980).
- [14] Gelbard MK, Walsh R, Kaufman JJ. Collagenase for Peyronie's disease experimental studies. *Urol Res* 10:135–40 (1982).
- [15] Nagase H, Woessner JF. Matrix Metalloproteinases. *J Biol Chem* 274:21491–4 (1999). doi:10.1074/jbc.274.31.21491.
- [16] Borth W. Alpha 2-macroglobulin, a multifunctional binding protein with targeting characteristics. *FASEB J* 6:3345–53 (1992).
- [17] Postlethwaite, A.E., Lachman, L.B., Mainardi, C.L., Kang, A.H. Interleukin 1 stimulation of collagenase production by cultured fibroblasts. *J Exp Med* 157:801–6 (1983).
- [18] Raykha CN, Crawford JD, Burry AF, Drosdowech DS, Faber KJ, Gan BS, et al. IGF2 expression and  $\beta$ -catenin levels are increased in Frozen Shoulder Syndrome. *Clin Invest Med* 37:E262–7 (2014).
- [19] Stanton H, Melrose J, Little CB, Fosang AJ. Proteoglycan degradation by the ADAMTS family of proteinases. *Biochim Biophys Acta* 1812:1616–29 (2011). doi:10.1016/j.bbadis.2011.08.009.
- [20] Wilkinson JM, Davidson RK, Swingler TE, Jones ER, Corps AN, Johnston P, et al. MMP-14 and MMP-2 are key metalloproteinases in Dupuytren's disease fibroblast-mediated contraction. *Biochim Biophys Acta* 1822:897–905 (2012). doi:10.1016/j.bbadis.2012.02.001.
- [21] Krause C, Kloen P, ten Dijke P. Elevated transforming growth factor  $\beta$  and mitogen-activated protein kinase pathways mediate fibrotic traits of Dupuytren's disease fibroblasts. *Fibrog Tiss Rep* 4:14 (2011). doi:10.1186/1755-1536-4-14.
- [22] Koźma EM, Olczyk K, Wisowski G, Głowacki A, Bobiński R. Alterations in the extracellular matrix proteoglycan profile in Dupuytren's contracture affect the palmar fascia. *J Biochem* 137:463–76 (2005). doi:10.1093/jb/mvi054.
- [23] Beare AHM, O'Kane S, Krane SM, Ferguson MWJ. Severely impaired wound healing in the collagenase-resistant mouse. *J Invest Dermatol* 120:153–63 (2003). doi:10.1046/j.1523-1747.2003.12019.x.
- [24] Melling M, Karimian-Teherani D, Mostler S, Behnam M, Sobal G, Menzel EJ. Changes of Biochemical and Biomechanical Properties in Dupuytren Disease. *Arch Path Lab Med* 124:1275–81 (2000). doi:10.1043/0003-9985(2000)124<1275:COBABP>2.0.CO;2.
- [25] Thiruvoth F, Mohapatra D, Sivakumar D, Chittoria R, Nandhagopal V. Current concepts in the physiology of adult wound healing. *Plastic and Aesthetic Research* 2:250 (2015). doi:10.4103/2347-9264.158851.
- [26] Grinnell F, Zhu M, Parks WC. Collagenase-1 complexes with alpha2-macroglobulin in the acute and chronic wound environments. *J Invest Dermatol* 110:771–6 (1998). doi:10.1046/j.1523-1747.1998.00192.x.
- [27] Ruddell RG, Oakley F, Hussain Z, Yeung I, Bryan-Lluka LJ, Ramm GA, et al. A Role for Serotonin (5-HT) in Hepatic Stellate Cell Function and Liver Fibrosis. *Am J Pathol* 169:861–76 (2006). doi:10.2353/ajpath.2006.050767.
- [28] de Almeida Prado PS, Soares MF, Lima FO, Schor N, Teixeira VP. Amitriptyline aggravates the fibrosis process in a rat model of infravesical obstruction. *Int J Exp Pathol* 93:218–24 (2012). doi:10.1111/j.1365-2613.2012.00813.x.
- [29] Murphy AM, Wong AL, Bezuhly M. Modulation of angiotensin II signaling in the prevention of fibrosis. *Fibrog Tiss Rep* 8:7 (2015). doi:10.1186/s13069-015-0023-z.
- [30] Drummond AH, Beckett P, Brown PD, Bone EA, Davidson AH, Galloway WA, et al. Preclinical and clinical studies of MMP inhibitors in cancer. *Ann N Y Acad Sci* 878:228–35 (1999).

- [31] Fan H, Jiang W, Li H, Fang M, Xu Y, Zheng J. MMP-1/2 and TIMP-1/2 expression levels, and the levels of collagenous and elastic fibers correlate with disease progression in a hamster model of tongue cancer. *Oncol Lett* 11:63–8 (2016). doi:10.3892/ol.2015.3837.
- [32] Dolmans GH, Werker PM, Hennies HC, Furniss D, Festen EA, Franke L, et al. Wnt Signaling and Dupuytren's Disease. *N Engl J Med* 365:307–17 (2011). doi:10.1056/NEJMoa1101029.
- [33] Hinz B, Phan SH, Thannickal VJ, Prunotto M, Desmoulière A, Varga J, et al. Recent Developments in Myofibroblast Biology. *Am J Pathol* 180:1340–55 (2012). doi:10.1016/j.ajpath.2012.02.004.
- [34] Hutchinson JW, Tierney GM, Parsons SL, Davis TR. Dupuytren's disease and frozen shoulder induced by treatment with a matrix metalloproteinase inhibitor. *J Bone Joint Surg Br* 80:907–8 (1998).
- [35] Felici N, Marcoccio I, Giunta R, Haerle M, Leclercq C, Pajardi G, et al. Dupuytren Contracture Recurrence Project: Reaching Consensus on a Definition of Recurrence. *Handchirur Mikrochirur Plast Chir* 46(6):350-4 (2014). doi:10.1055/s-0034-1394420.
- [36] Satish L, Palmer B, Liu F, Papatheodorou L, Rigatti L, Baratz ME, et al. Developing an animal model of Dupuytren's disease by orthotopic transplantation of human fibroblasts into athymic rat. *BMC Musculoskelet Disord* 16:138 (2015). doi:10.1186/s12891-015-0597-z.
- [37] Hovius SER, Kan HJ, Verhoekx JSN, Khouri RK. Percutaneous Aponeurotomy and Lipofilling (PALF): A Regenerative Approach to Dupuytren Contracture. *Clin Plas Surg* 42:375–81 (2015). doi:10.1016/j.cps.2015.03.006.